

not be covalently attached to a receptor and it does not require the addition of a detergent to enhance fluorescence intensity.

Received: June 5, 2002 [Z19477]

- [1] a) A. M. Thompson, *Science* **1992**, 256, 1157–1165; b) J. M. Anglada, Ph. Aplincourt, J. M. Bofill, D. Cremer, *ChemPhysChem* **2002**, 3, 215–221.
- [2] D. Price, P. J. Worsfold, R. Mantoura, C. Fauzi, *Trends Anal. Chem.* **1992**, 11, 379–384.
- [3] a) S. Wilson, *Chem. Ind. (London)* **1994**, 7, 255–258; b) C. Hachem, F. Bocquillon, O. Zahraa, M. Bouchy, *Dyes Pigm.* **2001**, 49, 117–125; c) *Fed. Regist.* **2000**, 65, 75174–75179.
- [4] J. Chen, W. H. Rulkens, H. Bruning, *Water Sci. Technol.* **1997**, 35, 231–238.
- [5] *The Enzyme Handbook* (Ed.: T. E. Barman) Springer, Berlin, **1974**.
- [6] *Methods of Enzymatic Analysis* (Ed.: H. G. Bergmeyer), Verlag Chemie, Weinheim, **1984**.
- [7] D. B. Papkovsky, T. C. O'Riordan, G. G. Guilbault, *Anal. Chem.* **1999**, 71, 1568–1573.
- [8] a) P. Leanderson, K. Wennerberg, C. Tagesson, *Cancerogenesis* **1994**, 15, 137–139; b) A. Barbouti, P. T. Doulias, B. Z. Zhu, B. Frei, D. Galaris, *Free Radical Biol. Med.* **2001**, 31, 490–498.
- [9] D. A. Stavreva, T. Gichner, *Mutat. Res.* **2002**, 514, 147–152.
- [10] *Bioluminescence and Chemiluminescence* (Eds.: M. A. DeLuca, W. D. McElroy), Academic Press, New York, **1981**.
- [11] K. A. Fährnich, M. Prawda, G. G. Guilbault, *Talanta* **2001**, 54, 531–559.
- [12] H. Perschke, E. Broda, *Nature* **1961**, 190, 257–258.
- [13] a) J. Meyer, U. Karst, *Nachr. Chem. Tech. Lab.* **1999**, 47, 1116–1119; b) J. Weber, U. Karst, *Analyst* **2000**, 125, 1537–1538 and references therein.
- [14] L. M. Hirschy, T. F. van Geel, J. D. Winefordner, R. N. Kelly, S. G. Schulman, *Anal. Chim. Acta* **1984**, 166, 207–219.
- [15] Reagent: A solution containing $\text{EuCl}_3 \cdot 6\text{H}_2\text{O}$ (9.6 mg) and tetracycline hydrochloride (4 mg) in a MOPS buffer (200 mL, 10 mmol L^{-1}) of pH 6.9. The absorbance at 405 nm is $\approx 0.78 \pm 0.01 \text{ cm}^{-1}$. The reagent is stable for at least 1 month if stored at 4°C in the dark.
- [16] M. Mathew, P. Balaram, *J. Inorg. Biochem.* **1980**, 13, 339–342.
- [17] a) L. C. Thompson in *Handbook on the Physics and Chemistry of Rare Earths* (Eds.: K. A. Gschneidner, L. Eyring), North-Holland Publ., Amsterdam, **1979**, pp. 210–290; b) F. S. Richardson, *Chem. Rev.* **1982**, 82, 541–552; b) S. I. Klink, H. Keizer, F. C. J. M. van Veggel, *Angew. Chem* **2000**, 112, 4489–4491; *Angew. Chem. Int. Ed.* **2000**, 39, 4319–4321.
- [18] a) B. Alpha, J. M. Lehn, G. Mathis, *Angew. Chem.* **1987**, 99, 259–261; *Angew. Chem. Int. Ed. Engl.* **1987**, 26, 266–267; b) M. Xiao, P. Selvin, *J. Am. Chem. Soc.* **2001**, 123, 7067–7073.
- [19] Assay protocol for H_2O_2 : The aqueous sample (1 mL, containing 1 to 2 $\mu\text{g mL}^{-1}$ of H_2O_2) was added to the $[\text{Eu}(\text{tc})]^{15}$ (1 mL) in a cuvette, and the increase in luminescence intensity at $> 616 \text{ nm}$ was measured after 10 min (excitation at 405 nm). If a time-resolving reader was available, the lag time was adjusted to $> 30 \mu\text{s}$ and the integration time to 100 μs .
- [20] GOx assay procedure: The following solutions were placed in the wells of a microtiterplate: a) A $[\text{Eu}(\text{tc})]$ solution (in the ten-fold concentration of that given in [15], except for the buffer concentration); b) a solution of GOx (of unknown activity) in the same buffer; c) enough buffer to fill up to a volume of 200 μL ; d) β -D-glucose (50 μL , 28 mmol L^{-1}) in buffer, to start the reaction. The increase in fluorescence intensity between 610 and 630 nm was measured over 10 min ($\Delta I_{10\text{min}}$). The activity of GOx was calculated by use of a calibration graph, established by plotting $\Delta I_{10\text{min}}$ versus known activities of GOx. Note: citrate and phosphate interfere.
- [21] Procedure for glucose assay: The $[\text{Eu}(\text{tc})]$ reagent solution^[15] (1 mL) was placed in a cuvette, to which a solution of GOx (2 mL, 15 units, from Sigma, product no. G 2133) in MOPS buffer (10 mL) was added. The reaction was initiated by adding 2 mL of a sample containing between 2 and 10 mmol L^{-1} of glucose. The increase in luminescence intensity after 10 min ($\Delta I_{10\text{min}}$) at 616 nm (excitation at 400–440 nm)

was measured, and $\Delta I_{10\text{min}}$ used to read off the glucose concentration using a calibration graph established with β -D-glucose. Note: citrate and phosphate interfere.

- [22] G. Liebsch, I. Klimant, B. Frank, G. Holst, O. S. Wolfbeis, *Appl. Spectrosc.* **2000**, 54, 548–559.
- [23] We used a Genios Plus microplate fluorometer from Tecan AG, Grödig, Austria.
- [24] German Patent Application, No. 101 11 392.7 (9 March 2001).
- [25] a) R. A. Evangelista, A. Pollak, E. F. G. Gudgin Templeton, *Anal. Biochem.* **1991**, 197, 213–221; b) M. H. V. Werts, J. W. Hoofstraat, F. A. J. Geurts, J. W. Verhoven, *Chem. Phys. Lett.* **1997**, 276, 196–201; c) B. Alpha-Bazin, G. Mathis, *Nucleosides Nucleotides* **1999**, 18, 1277–1278.
- [26] F. van de Rijke, H. Zijlmans, S. Li, T. Vail, A. K. Raap, K. Anton, R. S. Niebala, H. J. Tanke, *Nature Biotechnol.* **2001**, 19, 273–275.

Conversion of *arachno*-Nonaborane into Azanonaborane: Unexpected Loss of a Firmly Integrated Boron Atom**

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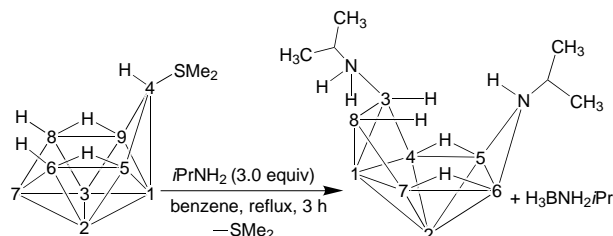
The polyhedral azanonaborane $(\text{RH}_2\text{N})\text{B}_8\text{H}_{11}\text{NHR}$ ($\text{R} = i\text{Pr}$, **1**) is easily prepared by treating dimethylsulfido *arachno*-nonaborane $4-(\text{Me}_2\text{S})\text{B}_9\text{H}_{13}$ with three equivalents of a primary amine.^[1] The reaction has been shown to proceed stepwise, by an initial ligand exchange to give $4-(\text{RH}_2\text{N})\text{B}_9\text{H}_{13}$, which reacts with an additional amine NH_2R^1 to give the mixed species $\text{R}^1\text{H}_2\text{NB}_8\text{H}_{11}\text{NHR}$.^[1b] These compounds have been shown to constitute a good entry into azacarbaborane^[2] and azametallaborane chemistry^[3] and may also be useful in neutron capture therapy.^[4] The transformation of $(\text{Me}_2\text{S})\text{B}_9\text{H}_{13}$ to $(\text{RH}_2\text{N})\text{B}_8\text{H}_{11}\text{NHR}$ involves the loss of one boron atom and cluster rearrangement. Herein we report the conversion of boron-substituted nonaboranes into azanonaboranes. These experiments make it possible to determine which boron atom is eliminated, and to speculate on the mechanism of the cluster rearrangement.

A variety of B-substituted $\text{B}_{10}\text{H}_{14}$ derivatives are known.^[5] These can be converted readily by a two-step process via 6,9- $(\text{Me}_2\text{S})_2\text{B}_{10}\text{H}_{12}$ derivatives into the corresponding *arachno*-nonaborane system.^[6] We prepared some ethyl, bromine, and deuterio derivatives of decaborane(14), which are stable under the reaction conditions (neither the bromine atom nor the ethyl group can be removed by Et_3N ,^[6a] and no deuterium exchange has been noted on heating the tetradeuterated $(\text{Me}_2\text{S})\text{B}_9\text{H}_{13}$ with diethylamine under reflux in benzene^[6c]).

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[**] We thank the Deutsche Forschungsgemeinschaft for support.

The following $(\text{Me}_2\text{S})\text{B}_9\text{H}_{13}$ compounds with labeled B-atoms were prepared (Scheme 1, Table 1): Et at B² (**2**) or B⁷ (**3**), Br at B² (**4**) or B⁶ (**5**) or B¹ (**6**), D at B¹, B², B³, and B⁷ (**7**). The presence and the positions of the substituents were confirmed by NMR and IR spectroscopy and mass spectrometry.



Scheme 1. Conversion of the B₉ clusters **2–7** into the B₈N clusters **1, 8–12**. The numbering of the clusters is given in Table 1, (*exo*-H atoms are omitted for clarity).

Table 1. Position of the substituents on the B₉ and B₈N clusters (see Scheme 1).

B ₉	Substituent	Position on B ₉ cluster	Position on B ₈ N cluster	B ₈ N
2	Et	2	2	8
3	Et	7	6	9
4	Br	2	2	10
5	Br	6	7	11
6	Br	1	–	1
7	D	1, 2, 3, 7	2, 5, 7 or 2, 4, 6	12

The labels of the B₉ cluster appeared in the following positions of the B₈N cluster after reaction with isopropylamine (Scheme 1, Table 1): Et at B² of **2** ⇒ B² in **8**; Et at B⁷ of **3** ⇒ B⁶ in **9**; Br of B² in **4** ⇒ B² in **10**, and the Br of B⁶ in **5** ⇒ B⁷ **11**, the Br of B¹ in **6** ⇒ bromine free B₈N cluster **1**. The 1,2,3,7-tetradeuterated nonaborane **7** was transformed to either the 2,4,6- or the 2,5,7- trideuterated B₈N cluster **12**. The NMR

spectroscopy results show clearly the loss of one deuterium atom (Table 2) while it was not possible to determine whether the remaining three deuterium atoms were at B², B⁵, and B⁷, or at B², B⁴, and B⁶.

Thus, the boron atoms B², B³, B⁶, and B⁷ in the B₉ cluster end up at the positions B², B⁴, B⁷, and B⁶ in B₈N, respectively, while B¹ of the B₉ cluster is lost during the conversion (Scheme 1).

According to these results we suggest the following stepwise mechanism for the conversion starting with the amino-substituted B₉ cluster (Scheme 2):

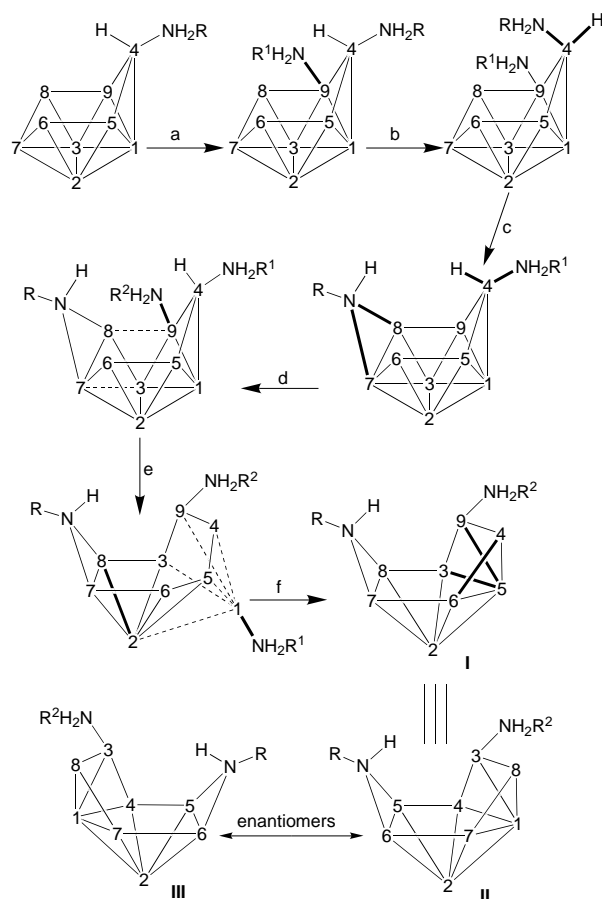
- Initially, an R¹NH₂ molecule attacks the 4-(RH₂N)B₉H₁₃ cluster at B⁹ or B⁵. This proposal is supported by two observations: 1) A 5-(RH₂N)B₉H₁₃ intermediate can be isolated from the reaction of 4-(Me₂S)B₉H₁₃ with RNH₂.^[7] 2) No reaction is observed when 5-OMe-4-(iPrH₂N)B₉H₁₂ is heated under reflux with excess iPrNH₂ in benzene.
- The RNH₂ moiety of B⁴ exchanges its position from *exo* to *endo* as described in the literature^[8] possibly aided by a weakened bond between B⁹ and B⁴.
- The RNH₂ moiety migrates from B⁴ to form a bridge between B⁷ and B⁸. This assumption is supported by the reaction rate of **3** (with Et on B⁷) which is slower than that of **2** (with Et on B²). In addition, the R¹NH₂ moiety migrates from B⁹ to the *exo* position of B⁴.^[7] (See also ref. [6b] for an analogous migration.)
- A third amine, R²NH₂, attacks at B⁹.
- The B⁸–B⁹ and B⁷–B³ bonds opens and a diamond-square-diamond (DSD) rearrangement^[9] which involves the atoms B², B³, B⁷, and B⁸ yields a bond between B² and B⁸.
- Migration of R¹NH₂ from B⁴ to B¹ leads to loss of the B¹–NH₂R¹ unit together with two additional H atoms. Specific elimination of B¹ was shown by reactions of the deuterium substituted **7** and the bromine substituted compound **6**. Closing the B⁵–B³, B⁵–B⁹, and B⁶–B⁴ connections completes the formation of the azanonaborane cluster.

The experimental basis for the proposed mechanism is the fate of the boron atoms B¹, B², B⁶, and B⁷ (on the basis of the

Table 2. ¹¹B and ¹H NMR spectroscopic data (CDCl₃ at 20 °C, 200 MHz) for B₈N clusters **1, 8–12**.^[a]

B ₈ N	B ¹	B ²	B ³	B ⁴	B ⁵	B ⁶	B ⁷	B ⁸	μ-H(4,5) μ-H(6,7)	NH
1	1.82 [2.57]	–55.61 [–0.65]	–21.46 [1.29]	–31.76 [0.86]	–11.11 [2.51]	–11.11 [2.51]	–33.41 [0.86]	–30.76 [0.55] [–0.64]	[–2.04] [–1.99]	[–1.56]
8	2.94 [2.58]	–43.91 [–]	–20.06 [1.28]	–32.3 [0.84]	–9.88 [2.34]	–10.6 [2.49]	–31.65 [0.84]	–30.62 [0.56] [–0.48]	[–1.9] [–1.75]	[–1.55]
9	0.76 [2.48]	–53.79 [–0.55]	–21.73 [1.26]	–31.67 [0.77]	–10.6 [2.21]	0.76 [–]	–32.91 [0.77]	–30.68 [0.52] [–0.55]	[–2.24] [–1.89]	[–1.36]
10	3.46 [3.06]	–41.77 [–]	–20.58 [1.21]	–28.8 [1.25]	–10.08 [2.75]	–10.08 [2.86]	–28.8 [1.25]	–28.8 [0.73] [–0.41]	[–1.34] [–1.34]	[–1.34]
11	12.06 [3.59]	–47.28 [–0.29]	–18.09 [1.18]	–28.77 [0.82]	–7.41 [2.70]	–4.72 [2.93]	–38.92 [–]	–23.25 [0.77] [–0.51]	[–2.47] [–1.72]	[–0.95]
12	1.64 [2.49]	–55.64 [–]	–21.46 [1.36]	–32.53 [0.74] ^[b]	–10.65 [–] ^[c]	–11.14 [2.55] ^[c]	–31.82 [–] ^[b]	–30.65 [0.46] [–0.61]	[–2.21] [–2.09]	[–1.64]

[a] All entries in ppm, δ(1H) in square parentheses. All spectra measured on a Bruker DP 200 spectrometer. [b], [c] A clear assignment of the deuterium substitutes to either B⁵ or B⁶ or to B⁴ or B⁷ is not possible.



Scheme 2. Mechanistic pathway of the conversion of the nonaborane cluster into the azanaborane cluster (**I** IUPAC numbering of the B_9 cluster, **II** and **III** IUPAC numbering of the B_8N cluster).^[10] Bold lines: new bonds; dashed lines: bonds to be broken.

experiments with the Et- and Br-substituted clusters) and B^3 (concluded from the experiments with a tetradeuterated cluster). Computed ^{11}B NMR chemical shifts, supported by 2D- ^{11}B COSY and 1H CW ^{11}B NMR spectra showed the Et group to be connected to B^6 , opposite to the *exo*-amino ligand at B^3 in the B_8N cluster **9**; the data further indicated that the Br atom at B^6 in the B_9 cluster **5** is located at B^7 and not at B^4 in the B_8N cluster **11**. The fate of the boron atoms B^5 , B^8 , and B^9 has not been clarified, but the proposed mechanism would require only a minimal rearrangement of the bonds (one DSD rearrangement and the closing of the cluster after the loss of B^1). The loss of B^1 , which is *not* part of the open face, is surprising. Quantum-mechanical computations might give indications about the feasibility of the proposed pathway, the relative stability of the proposed intermediates, the origin of the H atoms which leave together with B^1 , and the rearrangement of the other H atoms.

Experimental Section

1, 8–12: Isopropylamine (0.1 g, 1.74 mmol) was added to a solution of $(Me_2S)B_9H_{13}$ in dry benzene (10 mL, 0.1 g) at room temperature. The mixture was heated to reflux for 3 h. All volatile components were removed under vacuum and the resulting substance was recrystallized from ethanol:water (1:1). Compounds **8** and **9** were purified by using CH_2Cl_2 as eluent ($R_f=0.31$). For further purification the substance was

dissolved in $CHCl_3$:hexane (1:3) at $-20^\circ C$. The solution was filtered and the resulting filtrate was evaporated to dryness to yield the purified product. **1** (DCI): m/z (%) 214 (95) [M^+]; **8, 9** MS (EI, 750 eV, $200^\circ C$): m/z (%) 242 (24) [M^+]; **10, 11**, (EI, 750 eV, $200^\circ C$): m/z (%) 293 (18) [M^+]; **12** (FAB $^+$): m/z (%) 217 (100) [M^+].

Received: April 30, 2002
Revised: September 2, 2002 [Z19208]

- [1] B. M. Graybill, A. R. Pitochelli, M. F. Hawthorne, *Inorg. Chem.* **1962**, 1, 626–631; b) U. Dörfler, M. Thornton-Pett, J. D. Kennedy, *J. Chem. Soc. Dalton Trans.* **1997**, 2547–2550.
- [2] U. Dörfler, D. L. Ormsby, R. Greatrex, J. D. Kennedy, *Inorg. Chim. Acta.* **2000**, 304, 268–273.
- [3] a) U. Dörfler, J. D. Kennedy, L. Barton, C. M. Collins, N. P. Rath, *J. Chem. Soc. Dalton Trans.* **1997**, 707–708; b) U. Dörfler, P. A. Salter, X. L. R. Fontaine, N. N. Greenwood, J. D. Kennedy, M. Thornton-Pett, *Collect. Czech. Chem. Commun.* **1999**, 64, 947–958.
- [4] a) M. E. El-Zaria, U. Dörfler, D. Gabel, *J. Med. Chem.*, in press; b) C. Bauer, U. Dörfler, D. Gabel, *Eur. J. Med. Chem.*, in press.
- [5] a) N. J. Blay, I. Dunstan, R. L. Williams, *J. Chem. Soc.* **1960**, 430–433; b) J. A. Dopke, D. F. Gaines, *Inorg. Chem.* **1999**, 38, 4896–4897; c) R. F. Sprecher, B. E. Aufderheide, G. W. Luther III, J. C. Carles, *J. Chem. Soc.* **1974**, 96, 4404–4410.
- [6] a) T. L. Heying, C. Naar-Colin, *Inorg. Chem.* **1964**, 3, 282–285; b) H. Beall, D. F. Gaines, *Inorg. Chem.* **1998**, 37, 1420–1422; c) G. M. Bodner, F. R. Scholer, L. J. Todd, L. E. Senor, J. C. Carter, *Inorg. Chem.* **1971**, 10, 942–945.
- [7] L. F. K. Callaghan, U. Dörfler, D. T. McGrath, M. Thornton-Pett, D. J. Kennedy, *J. Organomet. Chem.* **1998**, 550, 441–444.
- [8] a) X. L. R. Fontaine, J. D. Kennedy, *J. Chem. Soc. Dalton Trans.* **1987**, 1573–1575; b) J. Müller, P. Paetzold, U. Englert, J. Runsink, *Chem. Ber.* **1992**, 125, 97–102.
- [9] R. Hoffmann, W. N. Lipscomb, *Inorg. Chem.* **1963**, 2, 231–232.
- [10] G. J. Leigh in *Nomenclature of Inorganic Chemistry*, Blackwell, Oxford, **1990**, pp. 217–225.

Fluorous Biphasic Catalysis without Perfluorinated Solvents: Application to Pd-Mediated Suzuki and Sonogashira Couplings**

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In catalytic reactions easy handling of the catalyst together with its straightforward recovery and possible reuse remain an important topic. A widespread solution to reach these goals is the application of immobilized catalysts. Immobilization can be achieved by covalent attachment to organic polymers or inorganic support materials.^[1] Alternatively, catalysts can be adsorbed on silica gel^[2–4] or on reversed-phase silica gel.^[5] In some cases the immobilization has a beneficial effect on the catalyst's stability.^[6,7] One profound advantage of such supported catalysts is the easy separation from the reaction

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[**] This work was supported by Fluka (Buchs, Switzerland).